MAHARSHI DAYANAND UNIVERSITY ROHTAK Department of Zoology Ph.D. course work 2020-21

PROGRAMME SPECIFIC OUTCOMES

PSO1: The Doctor of Philosophy program is designed to prepare each student to actively participate in research and teaching in the field of Zoology along with other fields of Life Sciences and in a University or a Research organization.

PSO2: Students are exposed to advanced experimental and theoretical techniques, encouraged to attend National and International conferences as well as workshops during the program.

PSO3: Several research areas of Zoology are interdisciplinary in nature and are funded by various funding agencies, giving students a flavour for both applied and basic research.

PSO4: Students in this programme acquire knowledge, critical thinking skills, and experience in conducting cutting-edge research. Students would gain proficiency in research methodology and assessment techniques in animal science.

PSO5: Students with a PhD degree either pursue a post-doctoral position aiming for an academic career or find employment in industrial R&D laboratories.

Duration: One Semester (Six months) **Total Credit requirement:** 14 credits **Program Structure: Ph.D. in Zoology**

SEMESTER 1						
Course Code	Nomenclature of Course	Theory marks (end semester examination)	Internal Assessment marks	Maximum marks	Hours /Week	Credits
20ZOOPH11C1 (Compulsory for all Ph.D. Course work)	Research Methodology	80	20*	100	4	4
20MPCC1 (Compulsory for all Ph.D. Course work)	Research and Publication Ethics	40	10**	50	2	2
20ZOOPH11C3	Biostatistics & Computers	80	20*	100	4	4
20ZOOPH11C4	Applications of Techniques in Animal Sciences	80	20*	100	4	4
Total marks/Credits				350		14

Ph.D. COURSE WORK

Scheme of Examination of Ph.D. (Course Work) Examination

Note: The compulsory course on 'Research and Publication Ethics' shall be offered by Ch. Ranbir Singh Institute of Social and Economic Change for all UTDs/Centres/Institutes passed vide Resolution No. 27 of the 271st meeting of EC held on 29.7.2020.

*<u>Internal Assessment:</u>

Two assignments of 5 marks each Two presentations of 5marks each **<u>Internal Assessment</u>: One assignment of 5 marks each One presentation of 5 marks each

Pass percentage will be 50% in each paper.

Ph.D. Course Work in Zoology syllabus

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Name of the Program	Ph.D. Course work in Zoology	Program Code	ZOOPH
Name of the Course	Research Methodology	Course Code	20ZOO11C1
Hours/Week	4	Credits	4
Max. Marks.	80	Time	3 Hours
Note: The examiner has	s to set a total of nine qu	estions (two from each u	init and one compulsory
question consisting of sl	nort answer from all units	. The candidate has to at	tempt one question each
from each unit along the	compulsory question (5 x	16 = 80 marks)	
Course Objectives:			
1. To know about the res	earch perspective in life so	ciences	
2. To understand the rese	earch hypothesis formulati	on and testing	
3. To understand the eler	nents of research methodo	ology	
4. To develop skills w.r.	t. Research article/papers	writing skills	
5. To understand signific	cance of scientific program	ns in Life Sciences	
Course Outcomes:			
CO1: Students should be	e able to identify the overa	all process of designing a r	research study from its
inception to its re	port.		
CO2: Students should k	now the primary character	istics of quantitative resea	rch and qualitative
research.			
CO3: Students should be	e able to identify a researc	h problem stated in a stud	у.
CO4: Students should be	e familiar with ethical issu	es in educational research	, including those issues
that arise in using q	uantitative and qualitative	research.	
	Uni	t - I	
Meaning of Research in Biological Sciences - Purpose, Characteristics and Types of Research - Process of Research -Formulation of objectives - Formulation of Hypotheses - Types of Hypotheses - Methods of testing Hypotheses - Research plan and its components - Methods of Research (Survey, Observation, case study, experimental, historical and comparative methods) - Difficulties in Biological research			
Unit - II			
Identification and formation of research problem (Hypothesis). Elements in research methodology:			
Research design (CRD, RBD, LSD). Scientific database: Science Direct and Pubmed.			
Unit - III			
Ethical, legal, social and scientific issues in Biological Research. A brief idea about the funding			
agencies such as DST, D	BT, ICMR, CSIR and UG	C. Role of IPR in Researc	h and Development.
Unit - IV			
Writing of Research Proposal, Report and Research Paper: Meaning and types - Stages in			
preparation			
Characteristics - Structure - Documentation: Footnotes and Bibliography - Editing the final draft-			
Evaluating the final draft- Checklist for the of a good proposal/report/research paper. Basic knowledge of organizing conferences symposic workshop whibition etc.			
References:			
References:			
Research Methodology- G.R. Basotia and K.K. Sharma.			

Research Methodology- C.H. Chaudhary, RBSA Publication

Name of the Program	Ph.D. Course work in	Program Code	ZOOPH	
	Zoology			
Name of the Course	Research and Publication ethics	Course Code	20MPCC1	
Hours/Week	2	Credits	2	
Max. Marks.	A. Marks.40Time3 Hours			
Note: The examiner ha	s to set a total of nine qu	estions (two from each u	init and one compulsory	
question consisting of s	hort answer from all units	. The candidate has to at	tempt one question each	
from each unit along the	compulsory question (5 x	8 = 40 marks)		
Course Objectives:				
1. To study the phi	losophy of ethics			
2. To study the sci	entific conduct of research			
3. To study the put	olication ethics			
4. To know about	various journal citation dat	abases		
5. To know the im	portance of quality publication	ations		
Course Outcomes:	4 4 1 4 11 4			
By completion of course	the student is able to			
1. Ethics in conduct	t of scientific research			
2. Know the scient	inc misconducts	nonaltics of placianism		
5. How to avoid pl	agiarism and what are the j	penalties of plagfarism		
4. Know the quant	y of research publications			
J. White research a	IIU IEVIEW articles.	4 T		
DHILOSODHV AND E		ι-1		
1 Introduction to r	hilosophy definition nati	ire and scope concept hr	anches	
2 Ethics: definition	n moral philosophy natur	e of moral judgments and	reactions	
SCIENTIFIC CONDU	CT	e of moral judgments and	reactions	
1 Ethics with respect to science and research				
2. Intellectual hone	2 Intellectual honesty and research integrity			
3. Scientific misconducts: Falsification, Fabrication, and Plagiarism (FFP)				
4. Redundant publ	4. Redundant publications: duplicate and overlapping publications, salami slicing			
5. Selective reporting and misrepresentation of data				
Unit - II				
PUBLICATION ETHICS				
1. Publication ethics: definition, introduction and importance				
2. Best practices / standards setting initiatives and guidelines: COPE, WAME, etc.				
3. Conflicts of interest				
4. Publication misconduct: definition, concept, problems that lead to unethical behavior and				
vice versa, types	;			
5. Violation of pub	5. Violation of publication ethics, authorship and contributorship			
6. Identification of	6. Identification of publication misconduct, complaints and appeals			
7. Predatory publis	hers and journals			

Ph.D. Course Work syllabus in Zoology

Unit - III

DATABASES AND RESEARCH METRICS

(A) Databases

- 1. Indexing databases
- 2. Citation databases: Web of Science, Scopus, etc.
- (B) Research Metrics
 - 1. Impact Factor of journal as per Journal Citation Report, SNIP, SIR, IPP, Cite Score
 - 2. Metrics: h-index, g index, i10 index, altmetrics

Unit - IV Practice

OPEN ACCESS PUBLISHING

- 1. Open access publications and initiatives
- 2. SHERPA/RoMEO online resource to check publisher copyright & self-archiving policies
- 3. Software tool to identify predatory publications developed by SPPU
- 4. Journal finder/journal suggestion tools viz. JANE, Elsevier Journal Finder, Springer Journal Suggested, etc.

PUBLICATION MISCONDUCT

(A) Group Discussions

- 1. Subject specific ethical issues, FFP, authorship
- 2. Conflicts of interest
- 3. Complaints and appeals: examples and fraud from India and abroad

(B) Software tools (2 hrs.) :Use of plagiarism software like Tumitin, Urkund and other open source software tools

References:

- 1. Bird, A. (2006). Philosophy of Science, Routledge
- 2. P. Chaddah (2018) Ethics in Competitive Research: Do not get scooped; do not get plagiarised.
- 3. Indian National Science Academy (INSA), Ethics in Science Education, Research and Governance (2019).
- 4. Beall, J (2012), Predatory publishers are corrupting open access. Nature, 489(7415), 179.
- 5. National Academy of Sciences, National Academy of Engineering and Institute of Medicine (2009). On being a Scientist: A guide to Responsible Conduct in Research, Third Edition, national Academic press.

Name of the Program	Ph.D. Course work in	Program Code	ZOOPH
	Zoology		
Name of the Course	Biostatistics &	Course Code	20ZOOPH11C3
	Computers		
Hours/Week	4	Credits	4
Max. Marks.	80	Time	3 Hours

Ph.D. Course Work in Zoology syllabus

Note: The examiner has to set a total of nine questions (two from each unit and one compulsory question consisting of short answer from all units. The candidate has to attempt one question each from each unit along the compulsory question ($5 \times 16 = 80$ marks)

Course Objectives:

1. Biostatistics will help to train the scholars in the skilled application of statistical methods to the solution of problems encountered in public health and medicine.

2. Biostatistics help the students in formulating the scientific questions to be answered, determine appropriate sampling techniques, coordinate data collection procedures, and conduct statistical analyses to answer those scientific questions.

3. Biostatisticians also play vital role in the preparation of research material for publication.

4. Show an awareness of what the major computer components are and how they act as system

5. Appreciate that computers need instructions to operate and acquire simple programming skills

6. To foster among students an interest and confidence in using computers; To encourage an understanding of the implications of computers in the modern world

Course Outcomes:

CO1: Students would gain knowledge about the assumptions, technique and applications of ANOVA

CO2: Students would be able to develop and test research ideas and apply the knowledge of research designs in planning and analysing research.

CO3: Students would gain knowledge about office applications of computer in research.

Unit - I

Variables in Biology, Collection, classification and tabulation of data. Frequency distribution, Diagrammatic and Graphical presentation of statistical data, Sampling techniques. Specific applications of measures of Central tendency, Dispersion, Skewness and Kurtosis in research.

Measures of Relationship: Correlation – Simple, Partial and multiple-Regression-Simple and multiple-Association of Attributes – applications in research.

Unit - II

PROBABILITY: - Meaning, Fundamental Concepts, Approaches to measurement of Probability, Random experiments, sample space, events. Mathematical definition of probability of an event. Use of permutations and combinations in calculation of probability.

PROBABILITY DISTRIBUTIONS: - Distribution of binomial, poisson and normal variables and their fittings only Binomial, Poisson and Normal, (areas method only) Distributions (including problems).

Unit - III

Hypothesis Testing and estimation: Fundamentals of hypothesis testing-Standard error point and interval estimates-Important non-parametric tests: Sign, Run Kruskal-Wallis tests and Mann – Whitney test. Level of significance. Definitions and applications of Chi-square test, 't' and 'f' test. Meaning of analysis of variance with linear models. Analysis of variance for one-way classified data, analysis of variance for two-way classified data.

Unit - IV

Computer Basics: Course introduction, MS Windows basics, UNIX basics, File management, E-mail (PINE, EUDORA, Internet mail), File Transfer (ftp, WSftp).

Office Applications: MS Office 2000/XP including MS Word, MS Excel, MS PowerPoint. **References:**

Elements of Biostatistics in Health Science- W. Daniell.

• Statistical Methods for Research: S. Singh et al (1988) Central Publishing Ludhiana.

- Fundamental of Statistics D. N. Enhance. ٠
- Statistical Methods: S.P. Gupta. S. Chand Publication Fundamentals of Biostatistics- Khan and Khanna, Ukaz Publication

Name of the Course Longy Course of Techniques Course Code 20ZOOPH11C4 Mours/Week 4 Credits 4 Max. Marks. 80 Time 3 Hours Note: The examiner has to set a total of nine questions (two from each unit and one compulsory question consisting of short answer from all units. The candidate has to attempt one question each from each unit along the compulsory question (5 x 16 = 80 marks) Course Objectives: 1. This paper aims to provide an introduction to various tools and techniques used to gain insight into biological samples and chemicals – life expectancy, precautions and their uses. 3. Solutions preparation, storage, stability, precautions, uses and their mechanism of action. 4. 4. The focus is on studying the techniques used for imaging, isolation, purification and characterization of bio-molecules etc – principles and applications in various areas of sciences. COI: Students would be able to develop basic appreciation of the underlying principles and practical strategy of the analytical and preparative techniques that are fundamental to study and understanding of ithe processes CO2: Students would be able to develop basic concepts and practical aspects of various kinds of Microscopy, Spectroscopy and separation techniques. CO3: Students would be able to inderstand the concept of radioisotope techniques, molecular biology techniques and their applications in research: types of microscopy; microtomy.	Name of the Program	Ph.D. Course work in Zoology	Program Code	ZOOPH
Techniques in Animal Sciences 4 Max. Marks. 80 Time 3 Hours Note: The examiner has to set a total of nine questions (two from each unit and one compulsory question consisting of short answer from all units. The candidate has to attempt one question each from each unit along the compulsory question (5 x 16 = 80 marks) Course Objectives: 1. This paper aims to provide an introduction to various tools and techniques used to gain insight into biological processes. 2. Handling of biological samples and chemicals – life expectancy, precautions and their uses. 3. Solutions preparation, storage, stability, precautions, uses and their mechanism of action. 4. The focus is on studying the techniques used for imaging, isolation, purification and characterization of bio-molecules etc – principles and applications in various areas of sciences. Course Outcomes: CO2: Students would be able to develop basic appreciation of the underlying principles and practical strategy of the analytical and preparative techniques that are fundamental to study and understanding of life processes CO3: Students would be able to understand the concept of radioisotope techniques, molecular biology techniques and their applications in research. Wint I Analysing the application of techniques in animal sciences research: types of microscopy, microtomy. Biophysical methods: Analysis of biomolecules using UV/visible, fluorescence, circular dichroism, MR and ESR spectroscopy, st	Name of the Course	Applications of	Course Code	20ZOOPH11C4
Animal Sciences Hours/Week 4 Max. Marks. 80 Time 3 Hours Note: The examiner has to set a total of nine questions (two from each unit and one compulsory question consisting of short answer from all units. The candidate has to attempt one question each from each unit along the compulsory question (5 x 16 = 80 marks) Course Objectives: 1 This paper aims to provide an introduction to various tools and techniques used to gain insight into biological samples and chemicals – life expectancy, precautions and their uses. 3. Solutions preparation, storage, stability, precautions, uses and their mechanism of action. 4. The focus is on studying the techniques used for imaging, isolation, purification and characterization of bio-molecules etc – principles and applications in various areas of sciences. Course Outcomes: COI: Students would be able to develop basic appreciation of the underlying principles and practical strategy of the analytical and preparative techniques that are fundamental to study and understanding of life processes CO2: Students would be able to develop basic concepts and practical aspects of various kinds of Microscopy, Spectroscopy and separation techniques. NMR and ESR spectroscopy, structure determination using X-ray diffraction and NMR; analysis using light scattering, different types of microscopy; microtomy, Biophysical methods: Analysis of biomolecules using UV/visible, fluorescence, circular dichroism, NMR and ESR spectroscopy, structure determination using X-ray diffraction and NMR; analysis using light scattering, different types of mass		Techniques in		
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Unit - IV	~	Uni	t - IV	
Computational methods: Nucleic acid and protein sequence databases; data mining methods for				
sequence analysis, web-based tools for sequence searches, motif analysis and presentation.				
Phylogenetic implications of computational data.				
tissues and cells, molecular imaging of radioactive material, safety guidelines: Immunoassavs &				

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diagnostic applications

References:

- Molecular cloning A Laboratory Manual 3rd edition Vol. 1, 2, 3- Sambrook and Russell, Churchill press, 2007
 Molecular Cell Biology, J. Darnell, H. Lodish and D. Baltimore Scientific American Book •